

DNA Methylation Bisulfite Kit

RK30203



Instruction Manual

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1. Introduction

The DNA Methylation Bisulfite Kit combines DNA denaturation and bisulfite conversion in one step and includes an additional heating step to promote rapid DNA denaturation. This product is a ready-to-use conversion reagent. Simply add this reagent to the DNA sample, and the conversion can be completed within 40 minutes. Subsequently, use the unique low-elution spin columns for desulfonation and purification of the converted DNA. The kit can accommodate conversion of samples ranging from 100 pg to 2 µg, with a conversion rate of unmethylated cytosine to uracil ≥99%. The high-yield converted DNA is highly suitable for applications such as PCR, qPCR and next-generation sequencing.

2. Components

Components	Component number	10 RXN	50 RXN	200 RXN
Conversion reagent	RM30451	1.3 mL	1.3 mL*5	1.3 mL*20
M-Binding buffer	RM30445	6 mL	30 mL	125 mL
M-Wash buffer	RM30446	1.2 mL	6 mL	24 mL
M-Desulphonation buffer	RM30447	2 mL	10 mL	40 mL
M-Elution buffer	RM30448	200 µL	1 mL	4 mL
Spin-column	RM30449	10 pk	50 pk	200 pk
Collection tube	RM30450	10 pk	50 pk	200 pk

3. Storage and Transport Conditions

Transport and store at room temperature.

4. Procedure

- ❑ Add 130 µl of Lightning Conversion Reagent to 20 µl of a DNA sample;



98°C, 8min;
54°C, 40min;
4°C, hold;

- ❑ Purification of Conversion Products;

30min;

4.1. Buffer Preparation

4.1.1. Before the first use, ethanol should be added to the M-Wash Buffer according to the instructions on the reagent bottle label. Check the box to mark it to avoid multiple additions. Tighten the bottle to prevent ethanol evaporation.

10 RXN: Add 4.8 mL of ethanol to 1.2 mL of M-Wash Buffer;

50 RXN: Add 24 mL of ethanol to 6 mL of M-Wash Buffer;

200 RXN: Add 96 mL of ethanol to 24 mL of M-Wash Buffer.

4.2. DNA denaturation and bisulfite conversion processes

4.2.1. In a PCR tube, add 130 μ l of Conversion Reagent to a 20 μ l of DNA sample. Mix well, then centrifuge quickly to ensure no droplets in the cap or sides of the tube.

Note: If the volume of DNA is less than 20 μ l, compensate with water. Samples >20 μ l must be processed using multiple conversion reactions. Replicate reactions can be cleaned using the same column.

4.2.2. Place the PCR tube in a thermal cycler and perform the following steps:

Temperature	Time
98°C	8 min
54°C	40 min
4°C	Hold

Note: Store at 4°C for up to 20 hours. The 4°C storage step is optional.

4.3. Purification

4.3.1. Add 600 μ l of M-Binding Buffer to a Spin-Column and place the column into a Collection Tube.

Note: The capacity of the collection tube with the column inserted is 800 μ l. Empty the collection tube whenever necessary to prevent contamination of the column contents by the flow-through.

4.3.2. Load the sample (from Step 4.2.2) into the Spin-Column containing the M-Binding Buffer. Close the cap and mix by inverting the column several times.

4.3.3. Centrifuge at full speed (12,000 x g) for 30 seconds. Discard the flow-through.

4.3.4. Add 100 μ l of M-Wash Buffer to the column. Centrifuge at full speed (12,000 x g) for 30 seconds.

4.3.5. Add 200 μ l of M-Desulphonation Buffer to the column and let stand at room temperature (20-30°C) for 15-20 minutes. After the incubation, centrifuge at full speed (12,000 x g) for 30 seconds.

4.3.6. Add 200 μ l of M-Wash Buffer to the column. Centrifuge at full speed (12,000 x g) for 30 seconds. Repeat this wash step.

4.3.7. Repeat step 4.3.6.

4.3.8. Discard the flow-through. Place the column into a 1.5 ml microcentrifuge tube and add 10 μ l of M-Elution Buffer directly to the column matrix. Centrifuge for 30 seconds at full speed (12,000 x g) to elute the DNA.

Note: Alternatively, water or TE (pH \geq 6.0) can be used for elution if required for your experiments. The DNA is ready for immediate analysis or store at or below -70°C for long term storage. The elution volume can be > 10 μ l depending on the requirements of your experiments, but small elution volumes will yield higher DNA concentrations.

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