

Product Components

Components	Component number	Sie-1	Size-2
		5 mL	25 mL
2X Universal SYBR Green Fast qPCR Mix *	RM21203	5 × 1 mL	25 × 1 mL

*Including ABclonal Hot Start Taq DNA polymerase, Mg²⁺, dNTPs, SYBR® Green I, Universal ROX Dye, etc.

Product Description

Real-time Quantitative PCR (qPCR) is a powerful technique in detecting initial DNA input in a PCR reaction by fluorescence signal accumulation. The DNA double strand bonded dye, SYBR® Green I is the most commonly used dye in qPCR. ABclonal 2X Universal SYBR Green Fast qPCR Mix contains de novel designed universal reference dye, which can realize higher signal resolution and suits for all currently used qPCR instruments (including high ROX mode, low ROX mode and No ROX mode required machines). It also contains Hot-Start Taq DNA polymerase to avoid unexpected amplification results.

Besides, ABclonal 2X Universal SYBR Green Fast qPCR Mix is an optimized qPCR reaction mix, It contains all required components in qPCR except primers and template. It is convenient for experiment and suitable for multiple species. The above features make it as an ideal experiment tool for gene quantitative research.

Instruments

No additional reference dye is required. ABclonal 2X Universal SYBR Green Fast qPCR Mix is suited for all currently used qPCR instruments (including high ROX mode, low ROX mode and No ROX mode required machine).

Products Storage

- 20°C for long term storage and 4°C for a short period and Light protection is demanded.

Materials Required

1. EP tubes, PCR tubes and other related materials.
2. qPCR specific primers and templates.
3. qPCR plates and seal membrane.

Usage Notes

1. Before using 2X Universal SYBR Green Fast qPCR Mix, please make sure that the mix is thawed completely and then placed it on ice for use.
2. Mix other qPCR components with Mix thoroughly and gently by pipetting or vortexing. After usage, place the mix back to -20°C for long time storage or 4°C for short period usage.
3. 2X Universal SYBR Green Fast qPCR Mix contains Taq polymerase, all operation should be performed on ice.
4. 2X Universal SYBR Green Fast qPCR Mix contains specially designed reference dye, which suits for all currently used qPCR machine. No extra ROX-like reference dyes should be added.
5. To avoid contamination, pipette tips with filters is suggested.
6. To guarantee better qPCR results, DNA template in good quality is suggested.

Protocol

Before Use

1. Specificity of primers should be checked and a final concentration of 0.2 μM is suitable for most of primers.
2. The length of amplification products is usually range from 70 bp to 200 bp.
3. Dilute the template in gradient.

4. Add 1 pg-50 ng DNA as PCR templates and a "No Template Control sample" is suggested.
5. To ensure the confidence of experiment, at least 2 repeats of each samples is suggested.

Experiment procedure

1. Prepare the following reaction systems on ice:

Components	Volume
2X Universal SYBR Green Fast qPCR Mix	10 μ L
Forward Primer (10 μ M)	0.4 μ L
Reverse Primer (10 μ M)	0.4 μ L
gDNA or cDNA (<50 ng)	2 μ L
ddH ₂ O	to 20 μ L

- (1) Dissolve 2X Universal SYBR Green Fast qPCR Mix at room temperature then placed it on ice for further usage. Before using, agitate the Mix thoroughly and centrifuge to collect all solution.
- (2) Calculate the amount of mix need, generally a 10% extra amount is suggested.
- (3) Dispense solution in sterile PCR or EP tubes in case of any contamination.
- (4) Add all components listed in above table, agitate the tubes gently to mix thoroughly (avoid bubbles) and centrifuge it.
- (5) Dispense the reaction solution into qPCR plates and seal the plates with optical membrane.
- (6) 2500 rpm centrifuge the qPCR plates to collect all solution.

2. Program qPCR reaction as follows:

Stage 1	Denaturation	Reps: 1	95 °C	3 min
Stage 2	Cycles	Reps: 40-45	95 °C	5 s
			60 °C	30-34 s *
Stage 3	Melt Curve	Reps: 1	default	

*Confirm there is a signal collection step after each extending step. The extending time is varied according to different machines: 30 s for StepOne Plus, 31 s for 7300 and 34 s for 7500.

Data Analysis

1. Draw a standard curve according to Ct values of endogenous gene. The value of R² should be more than 0.98 and the slope of curve should be in the range of -3 to -3.5 which means the PCR amplification efficiency is in the range of 90% to 120%.
2. The standard deviation (STD) of Ct values should be less than 0.2 and the variation of Ct value for different experiment should be less than 0.5 (the threshold value of different experiments should be same when comparing Ct values).
3. The single melt curve indicate the no non-specific amplification products or primer dimmers, and the T_m value in melt curve is usually in the range of 80 to 95°C.

Troubleshooting

Melt Curve Show Multiple Peaks

- a. Primer Design: Design the primer following basic primer design protocols.
- b. Primer Concentration Too High: lower down the concentration of primers.

Unusual Amplification Curves

- a. Amplification Curve Not Smooth: Too low amplification signal, increase the template input and make sure the qPCR Mix is stored properly.
- b. Inconsistent Amplification Curve: Bubbles causes abnormal qPCR results, centrifuge the plate prior to running it.
- c. Abnormal Amplification Curves: the default baseline value of machine is set to be from 3 to 15, the baseline setting can

be changed according actual amplification conditions. Besides, the degradation of template may affect the curve.

No Amplification Curves after Reaction

- Not Enough PCR Cycles: the PCR cycle number is usually set to be 40. It should be noted a higher cycle number may increase the background signal.
- Primer Degradation: Use electrophoresis to confirm the integrity of primers.
- Confirm the Signal Collection Step: the signal collection step are usually set to be after the annealing-extending step for two-step qPCR and after extending step for Three-step qPCR.
- Template Input Too Low: Increase template concentration or add extra repetition.
- Template Degradation: Use freshly prepared template (Use electrophoresis to confirm integrity of template).
- Not Enough Initial Denaturation Time: 2X Universal SYBR Green Fast qPCR Mix uses Hot-Start Taq polymerase, the pre-denaturation time should be at least 3 min.

Ct Value Too Late

- Low Amplification Efficiency: Optimize reaction condition or change primer.
- Template Input Too Low: Increase template concentration or add extra repeat.
- Template Degradation: Use freshly prepared template (Use electrophoresis to confirm Integrity of template).
- Too Long PCR Products: The length of amplification products is usually in the range of 70 bp-200 bp.
- PCR Inhabitation Reagent: use new template or dilute the template.
- Too Short Pre-denaturation Time: 2X Universal SYBR Green Fast qPCR Mix contains Hot-Start Taq polymerase, the pre-denaturation time should be at least 3 min.

NTC Shows Amplification

- Contamination: Use sterile water to conduct experiment and the all operation is suggested to be done in clean room to avoid aerosol contamination.
- Non-Specific PCR Products: analyze with melt curve.

Inconsistent Results

- Inconsistent Sample Added: Use proper pipetting techniques.
- Inconsistent Temperature in qPCR Machine: ensure periodic machine calibration.
- Template Concentration Too Low: the lower template input, the poorer qPCR result is. Increase the template concentration.
- Inconsistent Threshold Set: when comparing the qPCR results in different plates, make sure the threshold value of each experiments is same.

Related Products

Name	Catalog	Size
ABScript Neo RT Master Mix for qPCR	RK20432	10 RXN / 100 RXN
ABScript Neo RT Master Mix for qPCR with gDNA Remover	RK20433	10 RXN / 100 RXN
ABScript II cDNA First-Strand Synthesis Kit	RK20400	50 RXN / 100 RXN